Short Communication

Association of genetic variants of β-lactoglobulin gene with milk production in a herd and a superior family of Holstein cattle

Maryam Heidari¹, Mojtaba Ahani Azari²*, Saeed Hasani³, Alireza Khanahmadi⁴, Saeed Zerehdaran⁵

¹,²,³,⁵Department of Animal Sciences, Faculty of Agriculture, Gorgan University of Agricultural Sciences and Natural Resources, P.O. Box 49189-43464, Gorgan, I.R. Iran ⁴Department of Animal Sciences, Faculty of Agriculture, P.O. Box 163, Gonbad, I.R. Iran

Abstract
Polymorphism of the β-lactoglobulin (β-LG) gene in 101 cows belonging to the Holstein herd and a superior cow, producing more than 150 Kg milk/day, together with four offsprings was investigated by the Polymerase Chain Reaction-Restriction Fragment Length Polymorphism (PCR-RFLP) method. In the Holstein herd, the alleles A and B of the β-LG gene had frequencies of 0.53 and 0.47, respectively. The genotypes AA, AB and BB of the β-LG gene were estimated to have frequencies of 0.257, 0.544 and 0.198, respectively. Genotypes were distributed according to the Hardy-Weinberg equilibrium. Results indicated that the β-LG genotypes significantly affected (P< 0.01) milk yield (genotype AA being more effective than genotype BB). The superior cow and her progenies were all heterozygotes (AB).

Key words: β-LG; Holstein; Milk production; Polymorphism; PCR-RFLP

β-lactoglobulin (β-LG) is the major whey protein of ruminant species and is also present in the milk of many, but not all mammalian species (Kontopidis et al., 2004). β-LG is located on chromosome 11 in the cow (Daniela et al., 2008) and has 12 known variants in which A and B variants are the most frequent ones (Rachagani et al., 2006). The β-LG binding affinity for retinol (Frapin et al., 1993) and fatty acids (Fugate and Song, 1980) has been demonstrated, which suggests a possible role of β-LG in the transport and metabolism of these components (Lum et al., 1997; Eigel et al., 1984). There are many contradictory reports on the association between genetic variants of β-LG and milk production and composition (Tsiaras et al., 2005; Aleandri et al., 1990; Ng-Kawi-Hang et al., 1986; Ng-Kawi-Hang et al., 1984). Several researches have shown that the BB genotype of β-LG is associated with higher fat and increased cheese yields (Wedholm et al., 2006; Aleandri et al., 1990; Ng-Kawi-Hang et al., 1986). An association of β-LG BB with higher protein yield has also been reported (Ng-Kawi-Hang et al., 1984). On the contrary, Bovenhuis et al. (1992) reported lower protein yield for this genotype. In various studies, the AA genotype of the β-LG gene has been associated with higher milk production (Ikonen et al., 1999; Bovenhuis et al., 1992; Ng-Kwai-Hang et al., 1990) and higher milk protein (Strazalkowska et al., 2002; Ng-Kwai-Hang et al., 1990; Ng-Kwai-Hang et al., 1984). In other reports, the AB genotype has been associated with higher milk production (Tsiaras et al., 2005; Kaygisiz and Douan, 1999), lactose and protein yield (Tsiaras et al., 2005).

The current study was carried out from July 2008 to
November 2008 in the Molecular Genetics laboratory at the Department of Animal Sciences at Gorgan University of Agricultural Sciences and Natural Resources. The aim was to investigate $\beta$-LG polymorphism and its association with milk yield in a herd of Holstein cows. The blood samples were collected randomly from 101 Holstein cows (Behin Talise farm in Gorgan, Iran) having at least one lactation record. In the meantime, blood samples of a superior cow (producing more than 150 Kg milk/day) and her four female progenies from a farm near the region of the investigated herd were also collected. DNA was extracted, using the salting out extraction protocol (Miller et al., 1988). The 247 bp fragment, comprising a part of the IV exon and intron of the genomic DNA was amplified by using primers as suggested by Strazalkowska et al. (2002). The sequences of the primers were as follows: forward: 5’-TGT GCT GGA CAC CGA CTA CAA AAA G-3’; and reverse: 3’-GCT CCC GGT ATA TGA CCA CCC TCT-5’.

Polymerase Chain Reaction (PCR) was carried out, using the Personal Cycler™ thermocycler (Biometra, Germany) and the PCR Master Kit (CinnaGen Inc., Iran). The kit master mix consisted of 0.04 U/μl of Taq DNA polymerase, 10X PCR buffer, 3mM MgCl2 and 0.04 mM dNTPs (each). Each reaction mixture consisted of 12.5 μl of the master mix, 1 μl of each primer (5 pmol/μl) and some deionized water making up a final volume of 25 μl. The amplification program was as follows: initial denaturation at 94°C for 5 min followed by 35 cycles of denaturation at 94°C for 1 min, annealing at 60°C for 1 min, extension at 72°C for 1 min and a final extension at 72°C for 5 min. Digestion of PCR products was carried out, using the HaeIII restriction endonuclease (Fermentas, European Union) at 37°C for 16 h and then analyzed by electrophoresis on a 2.5% (w/v) agarose gel (Fig.1). Determination of gene and genotype frequencies and the $\chi^2$ test were carried out using the POPGene 32 software (Yeh et al., 1997).

The $\beta$-LG allele and genotype frequencies of the herd and those of other studies are summarized in

<table>
<thead>
<tr>
<th>Breeds</th>
<th>Allele Frequency A</th>
<th>B</th>
<th>Genotype Frequency AA</th>
<th>AB</th>
<th>BB</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Holstein</td>
<td>0.529</td>
<td>0.471</td>
<td>0.257</td>
<td>0.544</td>
<td>0.198</td>
<td>Current study</td>
</tr>
<tr>
<td></td>
<td>0.270</td>
<td>0.730</td>
<td>0.060</td>
<td>0.525</td>
<td>0.414</td>
<td>Celik, (2003)</td>
</tr>
<tr>
<td></td>
<td>0.516</td>
<td>0.484</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Kaygiziz and Douan, (1999)</td>
</tr>
<tr>
<td></td>
<td>0.420</td>
<td>0.580</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Sabour et al. (1993)</td>
</tr>
<tr>
<td></td>
<td>0.498</td>
<td>0.502</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Lunden et al. (1997)</td>
</tr>
<tr>
<td></td>
<td>0.520</td>
<td>0.480</td>
<td>0.284</td>
<td>0.471</td>
<td>0.245</td>
<td>Tsiaras et al. (2005)</td>
</tr>
<tr>
<td></td>
<td>0.231</td>
<td>0.769</td>
<td>0.037</td>
<td>0.387</td>
<td>0.576</td>
<td>Lin and Mcallister, (1986)</td>
</tr>
<tr>
<td></td>
<td>0.387</td>
<td>0.613</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Ng-Kawi-Hang and Kim, (1984)</td>
</tr>
<tr>
<td></td>
<td>0.520</td>
<td>0.480</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Ron et al. (1994)</td>
</tr>
<tr>
<td>Brown Swiss</td>
<td>0.440</td>
<td>0.560</td>
<td>0.172</td>
<td>0.289</td>
<td>0.537</td>
<td>Celik, (2003)</td>
</tr>
<tr>
<td>Sahiwal</td>
<td>0.170</td>
<td>0.830</td>
<td>0.031</td>
<td>0.276</td>
<td>0.693</td>
<td>Rachagani et al. (2006)</td>
</tr>
<tr>
<td>Tharparkar</td>
<td>0.390</td>
<td>0.610</td>
<td>0.023</td>
<td>0.733</td>
<td>0.244</td>
<td>Rachagani et al. (2006)</td>
</tr>
<tr>
<td>Guernsey</td>
<td>0.210</td>
<td>0.790</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Eenennaam and Medrano,(1991)</td>
</tr>
<tr>
<td>Milking Shorthorn</td>
<td>0.310</td>
<td>0.690</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Eenennaam and Medrano,(1991)</td>
</tr>
<tr>
<td>Red-sindhi</td>
<td>0.250</td>
<td>0.750</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Meignanalakshmi et al. (2001)</td>
</tr>
<tr>
<td>Ayrshire</td>
<td>0.158</td>
<td>0.842</td>
<td>0.190</td>
<td>0.279</td>
<td>0.702</td>
<td>Lin and Mcallister, (1986)</td>
</tr>
</tbody>
</table>

Figure 1. Electrophoresis of the digestion products of Haelll on 2.5% (w/v) agarose gel. Gel was stained with ethidium bromide, Lanes 1-8= digestion products of Haelll, Lane 1= BB genotype, Lanes 2, 4,5,7,9 =AB genotype, Lanes 3, 6= AA genotype, Lane M= Generuler™ 100-bp DNA Ladder Plus marker (Fermentas, European Union).

Table 1. $\beta$-LG gene and genotype frequencies in the Holstein breed and other breeds.
Table 1. In most of the studies, the frequency of the β-LG B allele was more than the A allele. The frequencies of the genetic variants A and B varied from 0.23 to 0.52 and 0.47 to 0.77, respectively in the Holstein breed. The results of the current study are in agreement with other studies, particularly with those of Ron et al. (1994) and Tsiaras et al. (2005). The maximum differences between A and B alleles’ frequencies have been observed in the Ayrshire (Lin and Mcallister, 1986) and Sahiwal (Rachagani et al., 2006) breeds. It must be noted that differences between allele frequencies of the gene depend on economic strategies and animal breeding programs for every breed and herd. Genotype of the superior cow and her progenies were determined as AB by using the same method.

A single-trait fixed model was used to examine the effect of β-LG genotypes on milk production. The mature equivalent milk yield records, corrected for missing data, were used. Statistical analysis was performed using the Generalized Least squares Means (GLM) procedure of the SAS software (2002) and comparison of the least squares means was carried out using the Tukey-Kramer test (SAS software, 2002). The following fixed model was used:

\[ y = Xb + e, \]
with \( E(y) = Xb \) and \( \text{var}(y) = V = \text{var}(e) \)

where:
- \( X \) = known incidence matrix relating observation in \( y \) to the fixed effects in \( b \),
- \( b \) = vector of levels of fixed effects,
- \( e \) = vector of random residual terms,
- \( E \) = the expectation of \( y \) and \( V \) = the variance of \( y \).

The fixed effects are represented by the calving years (2006, 2007 and 2008), seasons with 4 classes, parity with 8 classes and β-LG genotypes (AA, AB, BB).

Comparisons of the lactation milk yields of the β-LG genotypes showed that cows with the AA genotype produced more milk than animals with the BB genotype (\( P<0.006 \)) (Table 2). Comparison of expected and observed genotypic frequencies yielded a \( \chi^2 \) value of 0.78, suggesting that the β-LG locus was in Hardy-Weinberg equilibrium (\( P<0.05 \)).

The significantly higher milk production yield of cows with the AA genotype was in agreement with the results of other researches (Ikonen et al., 1999; Bovenhuis et al., 1992; Aleandri et al., 1990; Ng-Kuai-Hang et al., 1990).

Although there was a difference between AA and AB genotypes, but it was statistically not significant (Table 2). The superior cow, because of having both the A and B alleles and the AB genotypes, apparently was superior in milk production and fat yields, as well as the quality of cheese. Some reports have confirmed significant association between the AB genotype and higher milk yield (Tsiaras et al., 2005; Kaygisiz and Douan, 1999). Therefore, these genotypes (AA, AB) seem to be suitable candidates for selection aiming at improving milk production.

It must be pointed out that other factors might also affect milk yield of the cow, such as physiological status and other major genes. Hence, sequencings the genome of this cow will be an attractive theme for future studies to understand the main factors affecting this high production level.

Acknowledgments

The authors would like to thank Mr. Kabiri, owner of the Behin Talise farm, and Mr. Khari, owner of superior cow, for their kind help in providing blood samples. This research was financially supported by the University of Agricultural Sciences and Natural Resources, Gorgan, I.R. Iran.

References


